Synthesis and Biological Activity of Fluorinated Intermediates of the Methlonine Salvage Pathway

Michael E. Houston Jr.,^a David L. Vander Jagt,^b and John F. Honek^{a*}

^aDepartment of Chemistry, (GWC)², University of Waterloo, Waterloo, Ontario, Canada N2L 3G1

^bDepartment of Biochemistry, University of New Mexico, School of Medicine, Albuquerque, New Mexico

USA 87131

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Abstract: Fluorinated analogs of several of the intermediates of the methionine salvage pathway were synthesized and tested as inhibitors of the malarial parasite *Plasmodium falciparum* (FCB1, clone NC-1). Several compounds exhibit good activity against this chloroquine resistant strain. In addition, several of the analogs show antitumor activity.

The world wide incidence of malaria has been estimated to be 110 million clinical cases annually, with approximately 270 million humans infected and over one million malaria related deaths occurring each year. The development of *Plasmodium falciparum* strains resistant to chloroquine or to other antimalarial agents such as the dihydrofolate reductase inhibitor pyrimethamine has underscored the importance for the development of new agents against this protozoa.

The amino acid methionine's role in essential methylation reactions⁴ and polyamine biosynthesis⁵ is crucial to cellular viability. Methionine is the direct precursor to S-adenosylmethlonine (AdoMet) which is utilized in various cellular methylation and polyamine biosynthetic reactions (Figure 1). Salvage pathways exist for recycling S-adenosylhomocysteine (AdoHcy)⁶, a feedback inhibitor of various methylases, and 5'-deoxy-5'-methylthioadenosine⁷ (MTA) back into methionine. Depending on the organism methionine can be salvaged from MTA by two different pathways (Figure 1). Certain pathogenic protozoa and bacteria (*P. falcipanum, Klebsiella pneumoniae*) utilize a nucleosidase which converts MTA to 5-methylthioribose which is further transformed into 5-methylthioribose-1-phosphate by a specific kinase. However in mammals 5-methylthioribose-1-phosphate is directly produced from MTA by MTA phosphorylase. The completion of the pathway involves the multi-step conversion of 5-methylthioribose-1-phosphate into methionine and these individual steps have been investigated in some detail.⁸ The absence of the MTA nucleosidase and kinase in mammals has very recently been regarded as a possible difference between mammals and some protozoa worthy of drug development.⁹

Because of the unique steric and electronic properties of fluorine 10, our interest in the design of inhibitors of the above pathways currently involves the incorporation of fluorine into methionine and salvage pathway intermediates. 11 Therefore the corresponding fluorinated analogs at the methionine, nucleoside and ribose levels were synthesized and tested against a chloroquine resistant strain of *P. falciparum* (strain FCB1). We now wish to report our most recent results in this area including the good antimalarial and antitumor activity of several fluorinated analogs. Our approach was given added impetus by the recent reports by Riscoe and co-workers of the inhibitory activity of 5-trifluoromethylthioribose against a pyrimethamine resistant strain of *P. falciparum* 2 as well as *K. pneumoniae*. 13

Figure 1. Methionine Salvage Pathways

Our general synthetic approach to these compounds is applicable to both ribose and nucleoside analogs, several of which have been previously reported although the biological activity of these compounds have not been fully explored. The nucleoside analogs were prepared in the following manner (Scheme 1). Tosylation of 2', 3'-O-isopropylidene adenosine 14 followed by reaction with potassium thioacetate 15 gave rise to the thioacetate 1 Treatment of 1 with 4.5 equivalents of KOH in methanol/water 4:1 followed by alkylation with chlorodiffuoromethane at 60 °C gave the corresponding 5'-diffuoromethylthionucleoside 2 in 71 % yield. 16,17 The 5'-triffuoromethylthionucleoside 3 was prepared in 82 % yield by the reaction of 1 with lodotriffuoromethane in liquid ammonia under ultraviolet irradiation. A complete lack of reactivity of the adenine base under these fluoroalkylation reactions was observed. The fluorinated ethylthionucleosides were prepared by alkylation of thioacetate 1 with the corresponding fluorinated ethyl triflates in greater than 60 % yield 18. Removal of the isopropylidene protecting groups with 0.1 N HCl in dioxane resulted in the quantitative production of the desired 5'-diffuoromethylthioadenosine (7), 5'-trifluoromethylthioadenosine (8), 5'-diffuoroethylthioadenosine (9) and 5'-trifluoroethylthioadenosine (10). Due to the decreased stability of the monofluoroethylthionucleoside to deprotection steps, the desired compound 11 was prepared from the deprotected thioacetate 6.19

Chemical depurination of compounds **7** and **8** was achieved utilizing a modified procedure of Robins and Robins ²⁰ Acetylation with 10 equivalents of acetic anhydrade in pyridine followed directly by acetolysis with glacial acetic acid at 110 °C gave rise to the protected ribose analogs **12** and **13** in 53 % and 55 % respectively. Deprotection with

catalytic sodium methoxide yielded 5-difluoromethylthioribose (14) and 5-trifluoromethylthioribose (15).²¹ We have previously reported the synthesis of difluoromethylhomocysteine (16) and trifluoromethylhomocysteine (17).¹¹

Scheme 1. Synthesis of Fluorinated Methylthioadenosine and Methylthioribose Analogs

Reagents: (a) 4.5 eq. KOH, CF_2HCI , 60 °C, $MeOH/H_2O$ 4:1; (b) NH_3 (*f*), CF_3I , uv; (c) 2.1 eq. KOH, CF_2HCH_2OTf , MeOH; (e) 0.2 N HCI, $dioxane/H_2O$ 1:1; (f) 2.1 eq. KOH, CF_3CH_2OTf , MeOH; (e) 0.2 N HCI, $dioxane/H_2O$ 1:1; (f) 2.1 eq. KOH, CFH_2CH_2OTf , MeOH; (g) 10 eq. Ac_2O , pyridine; (h) 3 eq. Ac_2O , HOAc, 110 °C; (i) cat. NaOMe, MeOH.

The biological data for the fluorinated analogs is presented in Table 1. Because of the requirement of P. falciparum for exogenous methionine, 22 it was felt that fluorinated analogs of methionine may be transported into the malarial cell and cause cell death. However this was not observed with fluorinated analogs under the assay conditions utilized in this study. In addition, although trifluoromethylthioribose has been shown to inhibit the pyrimethamine resistant strain of P. falciparum (IC50 = 50 μ M)¹³, it is interestingly less active against the chloroquine resistant strain (IC50 > 100 μ M). A similar lack of activity of the difluoromethylthioribose analog against this strain of P. falciparum was also observed (IC50 > 100 μ M). The known transport and metabolism of MTA into human erythrocytes²³ and nucleosides into P. falciparum 22,24 indicated that fluorinated analogs of MTA could be cytotoxic to this protozoa. Indeed, the corresponding MTA

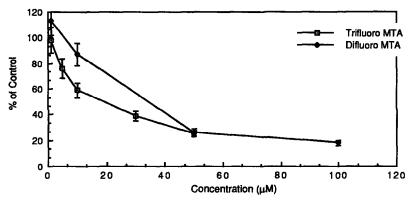
nucleoside analogs 7 and 8 are quite active against this strain (Figure 2). The fluorinated ethyl nucleosides were also found to be active (Table 1). Nucleoside transport may well be more efficient than transport of the ribose and methionine analogs and could account for the greater effectiveness of the nucleosides.

Table 1. Growth Inhibition of P. falciparum (FCB1)25

COMPOUND	<u>IС₅₀ (µМ)</u> *
L-Difluoromethylhomocysteine (16)	> 100
L-Trifluoromethylhomocysteine (17)	> 100
5-Difluoromethylthioribose (14)	> 100
5-Trifluoromethylthioribose (15)	>100
5'-Difluoromethylthioadenosine (7)	35
5'-Trifluoromethylthioadenosine (8)	18
5'-Monofluoroethylthioadenosine (11)	43
5'-Difluoroethylthioadenosine (9)	25
5'-Trifluoroethylthioadenosine (10)	85
* Triplicate determinations Values are / 10 9/ or loss	

^{*} Triplicate determinations. Values are +/- 10 % or less

Figure 2. Growth Inhibition of P. falciparum *



^{*} Triplicate values +/- S. D.

In addition, the biological activities of several of these analogs against various cancer cell lines 26 and HIV 27 were investigated. Difluoromethylhomocysteine (16) showed activity against small cell lung cancer DMS114 (GI $_{50}$ = 65 μ M, concentration of compound inhibiting cell growth by 50 %) and COLO 205 colon cancer (GI $_{50}$ = 92 μ M). Fluorinated nucleosides 7, 8 and 10 showed activity against non small cell lung cancer NCI-H522 with the difluoromethyl- and the trifluoroethylthionucleosides being more active (GI $_{50}$ = 86 μ M and 45 μ M respectively). These analogs showed no toxicity to other cell lines at the dose levels tested. Analogs 7, 8 and 10 were inactive against HIV infected T4 lymphocytes but showed toxicity (IC $_{50}$) to T4 lymphocytes of 222 μ M, 117 μ M and 245 μ M respectively

The novel nucleoside 8 shows the best inhibitory activity against *P. falciparum* (FCB1) of all fluorinated salvage pathway intermediates prepared to date. Investigations to determine the exact mechanism of action of these nucleosides is currently in progress; however, the biological data reported herein appears to indicate that the nucleosides may not be active due to their conversion into either fluorinated ribose or methionine level intermediates as both these analog types are devoid of activity. A more detailed investigation of the site(s) of action of these agents appears warranted.²⁸

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- Spectral data for nucleoside analogs: (7): ¹H NMR (CD₃OD, 250 MHz): 8.19 (s, 1H, purine), 8.15 (s, 1H, purine), 6.99 (t, 1H, CF₂H, J_HF = 56.9 Hz), 5.93 (d, 1H, H-1'), 4.84 (m, 1H, H-2'), 4.29 (m, 1H, H-3'), 4.19 (dt, 1H, H-4'), 3.21 (m, 2H, CH₂S); ¹³C NMR (DMSO): 155.9, 152.5, 149.5, 140.4, 122.0 (t, J_CF = 269.8 Hz), 119.1, 87.9, 83.7, 72.7, 72.5, 30.0. (8): ¹H NMR (CD₃OD): 8.19 (s, 1H, purine), 8.16 (s, 1H, purine), 5.94 (d, 1H, H-1'), 4.84 (m, 1H, H-2'), 4.33 (m, 1H, H-3'), 4.19 (m, 1H, H-4'), 3.33 (m, 2H, CH₂S); ¹³C NMR (CD₃OD): 151.5, 149.7,

145.8, 145.1, 132.3 (q, $J_{CF} = 305.1 \, Hz$), 120 5, 90.8, 84.4, 75.0, 73.9, 33.1. (9): ^{1}H NMR (CD₃OD): 8.18 (s, 1H, purine), 8.09 (s, 1H, purine), 5.92 (d, 1H, H-1'), 5.86 (tt, 1H, $\underline{CF_2HCH_2}$, $J_{HF} = 56.5 \, Hz$), 4.73 (t, 1H, H-2'), 4.27 (t, 1H, H-3'), 4.20 (m, 1H, H-4'), 2.93 (m, 2H, CH_2S), 2.81 (dt, 2H, CF_2HCH_2 , $J_{HF} = 16.7 \, Hz$); ^{13}C NMR (CD₃OD): 157.3, 153.9, 150.6, 141.5, 121.1, 117.9 (t, $J_{CF} = 240.9 \, Hz$), 90.3, 85.9, 74.7, 74.0, 36.1 (t, $J_{CF} = 240.0 \, Hz$), 35.9 (10): ^{1}H NMR (DMSO): 8 22 (s,1H, purine), 8.15 (s, 1H, purine), 5.93 (d, 1H, H-1'), 4.73 (m, 1H, H-2'), 4.36 (m, 1H, H-3'), 4.16 (m, 1H, H-4'), 2.95 (m, 4H, CF_3CH_2 , CH_2S); ^{13}C NMR (DMSO): 156.0, 152.5, 149.4, 140.2, 126.5 (q, $J_{CF} = 276.0 \, Hz$), 120.5, 90.7, 87.9, 83.8, 72.6, 35.1, 33.4 (q, $J_{CF} = 31.4 \, Hz$). (11): ^{1}H NMR (CD₃OD): 8.23 (s, 1H, purine), 8.13 (s, 1H, purine), 5.92 (d, 1H, H-1'), 4.71 (t, 1H, H-2'), 4.42 (dt, 2H, $J_{HF} = 47.4 \, Hz$), 4.25, (t, 1H, H-3'), 4.14 (m, 1H, H-4'), 2.93 (m, 2H, CH_2S), 2.75 (dt, 2H, $CFCH_2CH_2$, $J_{HF} = 21.0 \, Hz$). ^{13}C NMR (DMSO): 156.1, 152.6, 149.5, 139.8, 119.2, 85.7 (d, $J_{CF} = 217.8 \, Hz$), 82.1, 81.3, 72.5, 67.5, 34.2, 31.7 (d, $J_{CF} = 20.7 \, Hz$). In addition all compounds had mass spectra and infrared characteristics in complete agreement with the assigned structures.

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- Spectral data for ribose analogs. (14): 1 H NMR (CD₃OD): 7.14 (I, 1H, CF₂H α , J_{HF} = 57.3 Hz), 7.07 (I, 1H, CF₂H β , J_{HF} = 57.3 Hz), 5.21 (d, 1H, H-1 α), 5.08 (d, 1H, H-1 β), 4.14-3.79 (m, 3H, H-2, H-3, H4), 3.15-2.86 (m, 2H, CH₂S); 13 C NMR (DMSO): 122.9 (I, CF₂H α , J_{CF} = 269.0), 122.8 (I, CF₂H β , J_{CF} = 269.0 Hz),103.3 (C-1 β), 98.1 (C-1 α), 83.0 (C-4 α), 82.6 (C-4 β), 77.2 (C-2 β), 74.9 (C-3 β), 74.2 (C-3 α), 72.3 (C-2 α), 32.7 (C-5 β), 31.2 (C-5 α). (15): 11 H NMR (CD₃OD): 5.20 (d, 1H, H-1 α), 5.06 (d, 1H, H-1 β), 4.11-3.79 (m, 3H, H-2, H-3, H-4), 3.36 (m, 2H, CH₂S); 13 C NMR (CD₃OD): 132.7 (q, CF₃ α , J_{CF} = 304.2 Hz),132.6 (q, CF₃ β , J_{CF} = 304.2 Hz), 103.4 (C-1 β), 98.1 (C-1 α), 81.4 (C-4 α), 81.2 (C-4 β), 77.1 (C-2 β), 75.1 (C-3 β), 74.3 (C-3 α), 72.2 (C-2 α), 32.7 (C-5 β), 31.2 (C-5 α). 13 C assignments based on the assignments for α and β ribofuranoside obtained from; Bock, K. and Pedersen, C. *Adv. Carbohydr. Chem. Biochem.* 1983, 41, 27. In addition all compounds had mass spectra and infrared characteristics in complete agreement with the assigned structures
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